

Original Research

Two Simultaneously Occurring *Potamogeton* Species: Similarities and Differences in Seasonal Changes of Biomechanical Properties

Anna Maria Łoboda^{1*}, Robert Józef Bialik², Mikołaj Karpiński¹,
Łukasz Przyborowski¹

¹Institute of Geophysics, Polish Academy of Sciences, Warsaw, Poland

²Institute of Biochemistry and Biophysics, Polish Academy of Sciences, Warsaw, Poland

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Abstract

Two common *Potamogeton* species, *Potamogeton pectinatus* L. and *Potamogeton crispus* L., were collected in 2016 and 2017 from a lowland, sandy bed in the Wilga River in Poland to investigate the ability of the plants to adapt to changing hydrological conditions. Measurements included biomechanical properties as well as the morphological characteristics of their stems. Specifically, experiments included three-point bending and tension tests as well as stem diameter and cross-sectional morphology at various periods in the plants' life cycles. Detailed information about the seasonal changes in biomechanical traits and the similarities between the two investigated plants are presented. The data show significant differences in the three-point bending and tension parameters. The flexural rigidity proved to be the most sensitive parameter to changes in hydrological conditions during the season. The maximum forces in the three-point bending tests needed to complete the fracture of *P. crispus* were much higher, reaching values up to 0.097 N, than those for *P. pectinatus* (0.035 N), due to *P. crispus* having thicker shoots, which resulted in greater resistance to elastic deformity. Moreover, the modulus of elasticity values shows that *P. pectinatus* is much more prone to return to its original shape after the removal of the acting forces. For instance, the maximum Young's modulus for *P. pectinatus* was 116.23 MPa, whereas for *P. crispus*, the highest value was four times lower (26.60 MPa). The present study supplements an aquatic plant biomechanics database that has been created in recent years.

Keywords: curlyleaf pondweed, sago pondweed, mechanical traits, environmental management, aquatic macrophytes, bending, tension

*e-mail: aloboda@igf.edu.pl

Introduction

As stated by Nepf [1], the trade-off between floods and ecological management underlines the need for a reliable method to predict channel resistance in the presence of vegetation. The need for improving flow-vegetation-sediment interactions is important for engineering (e.g., hydraulic resistance estimation) [2-5] and ecological (e.g., stream restoration) [6-8] applications. Recently there has been an increased interest in management and control methods of invasive aquatic plants that have played an important role in river colonization during recent years [9-10 and literature therein]. Researchers have mostly concentrated on plant systematics or hybridization [e.g., 11] rather than on the basic biology, which should allow for precise parameterization of the interaction between aquatic plants and physical processes in rivers [12]. From this point of view, plant biomechanics seems to be an important property that should be regularly monitored, especially for invasive and phenoplastic aquatic plants, which have strong community dynamics. However,

not all freshwater species behave in the same way, and most of them – even those belonging to the same family – have completely different vegetative periods and reproductive methods that are often associated with their adaptation to seasonally changeable abiotic factors [13]. Many of these factors, including temperature or the availability of light and water nutrients, have been extensively studied [e.g., 14-15]. For instance, Riis et al. [14] claimed that aquatic macrophytes spread differently in relation to changing conditions (i.e., water temperature or light availability). In addition, as stated by de los Santos et al. [15], variations in plant biomechanics may be due to changes in water chemistry. However, only recently did Miler et al. [16] investigate the seasonal changes in the biomechanical properties of some aquatic plants (i.e., *Glyceria* spp., *Fontinalis* spp., *Ranunculus* spp., and *Myriophyllum* spp.). However, the authors did not find a general tendency in the seasonal changes of cross-sectional area, diameter, or biomechanical traits like ‘tension’ Young’s modulus or elasticity in the studied plants. They claimed that these

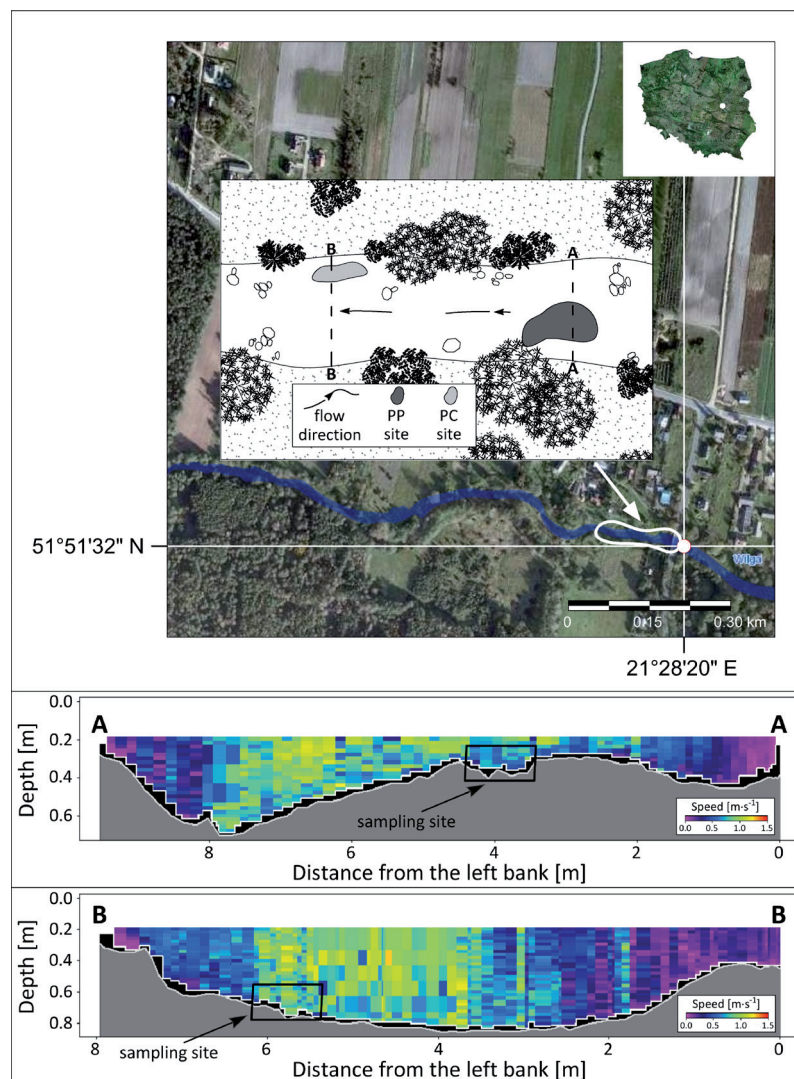


Fig. 1. Sampling site on the Wilga River with the velocity spatial distribution on channel sections of *P. pectinatus* (PP) and *P. crispus* (PC) measured using an acoustic Doppler current profiler.

plants did not use an increase in mechanical strength and/or stem flexibility as an adaptive strategy to resist harsh conditions during winter/spring.

This paper investigates the seasonal changes in biomechanical traits of two *Potamogeton* spp. that belong to a common submerged perennial herb [17]. These plants occur in rivers, lakes, and marshes around the world [18-19]. *P. pectinatus* is characterised by slender round shoots up to 3 m long with narrow linear leaves, whereas *P. crispus* has shorter heavily branched stems up to 2 m [17]. Both plants can reproduce vegetatively [18, 20]. The significant quality of these plants is a very successful survival strategy that contributes to their wide distribution [18, 21]. Moreover, the important dissimilarity that distinguishes *P. crispus* from other aquatic macrophytes is its unique life cycle. Firstly, the plants flower in spring, then die back or senesce in early summer. Finally, in autumn, the plants start to sprout again, given appropriate conditions in terms of light and water temperature [20]. Such phenological behaviour should be naturally considered with changes in biomechanical properties of these plants, which have only been investigated by a small number of researchers [16, 22-23] without any attention paid to the seasonal changes.

The first goal of this paper is to show detailed information about the seasonal changes in the biomechanical traits of the stems of two submerged aquatic plants, *P. pectinatus* and *P. crispus*, which live together in the Wilga River in Poland. The second goal is to supplement an aquatic plant biomechanics database that should have a strong impact on the ecological management of aquatic ecosystems. Moreover, we address two hypotheses:

1. The trends in changes in biomechanical properties of *Potamogeton* spp. differ between species at the same moment in phenological time due to the changes in their morphological structure and may be independent of their stage of growth.
2. The changes in hydrological conditions are closely related to changes in biomechanical parameters of *Potamogeton* spp., and they are independent of the plant species.

Materials and Methods

Sampling Sites and Data Collection

The studied plants, *P. pectinatus* and *P. crispus*, were collected in 2016 and 2017 over the course of the entire plant growth season. The sampling site (51°51'30" N, 21°28'20" E, Fig. 1) is located 60 km south of Warsaw, Poland, on the Wilga River. It is the right-bank tributary of the Vistula River, in which the channel bed consists of small stones and gravel with sand [24]. This location was chosen due to a common occurrence of two species from the family *Potamogetonaceae* with contrasting hydrological characteristics of plant locations in the river.



Fig. 2. The Wilga River with the view at the *P. pectinatus* sampling site.

In 2016 *P. pectinatus* was collected six times from May to October. The vegetation season in 2017 started in June and individuals were collected until November. After these periods, there were no visible macroscopic living specimens in the river, or the plants were damaged enough that they were not suitable for biomechanical measurements. The plants covered the greater part of the riverbed (Fig. 2), and samples were collected from the middle section of the channel, where the flow was around 0.5-0.7 m·s⁻¹ (Fig. 1). The sediment consisted of medium, moderately well sorted sand ($D_{16} = 0.29$ mm, $D_{50} = 0.46$ mm, $D_{84} = 1.97$ mm, Fig. 3). The second investigated species, *P. crispus*, was found and collected eight times in 2016 from May to December. In 2017 the plant was found and collected at the same sampling site only twice, in August and November. The habitat of this hydrophyte is located near the right river bank, covered by medium, poorly sorted sand ($D_{16} = 0.28$ mm, $D_{50} = 0.45$ mm, $D_{84} = 15.66$ mm; Fig. 3), where the flow is distinctly higher (1.0 m·s⁻¹; Fig. 1) than at the collection site of *P. pectinatus*. The high velocities and water discharge and the accompanying high water levels in the habitat of this plant made it impossible to collect this hydrophyte in May and October 2017 (Fig. 4).

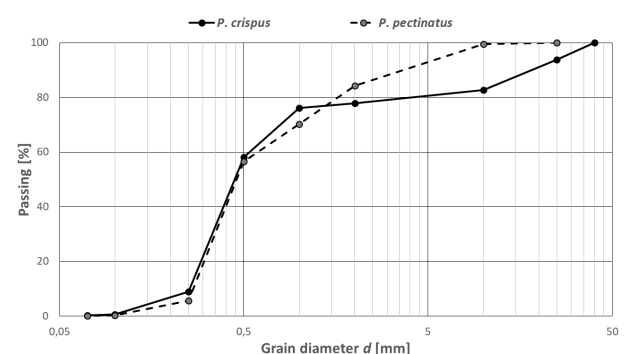


Fig. 3. Granulometric distribution curves of bed sediment from sampling sites of *P. pectinatus* and *P. crispus*.

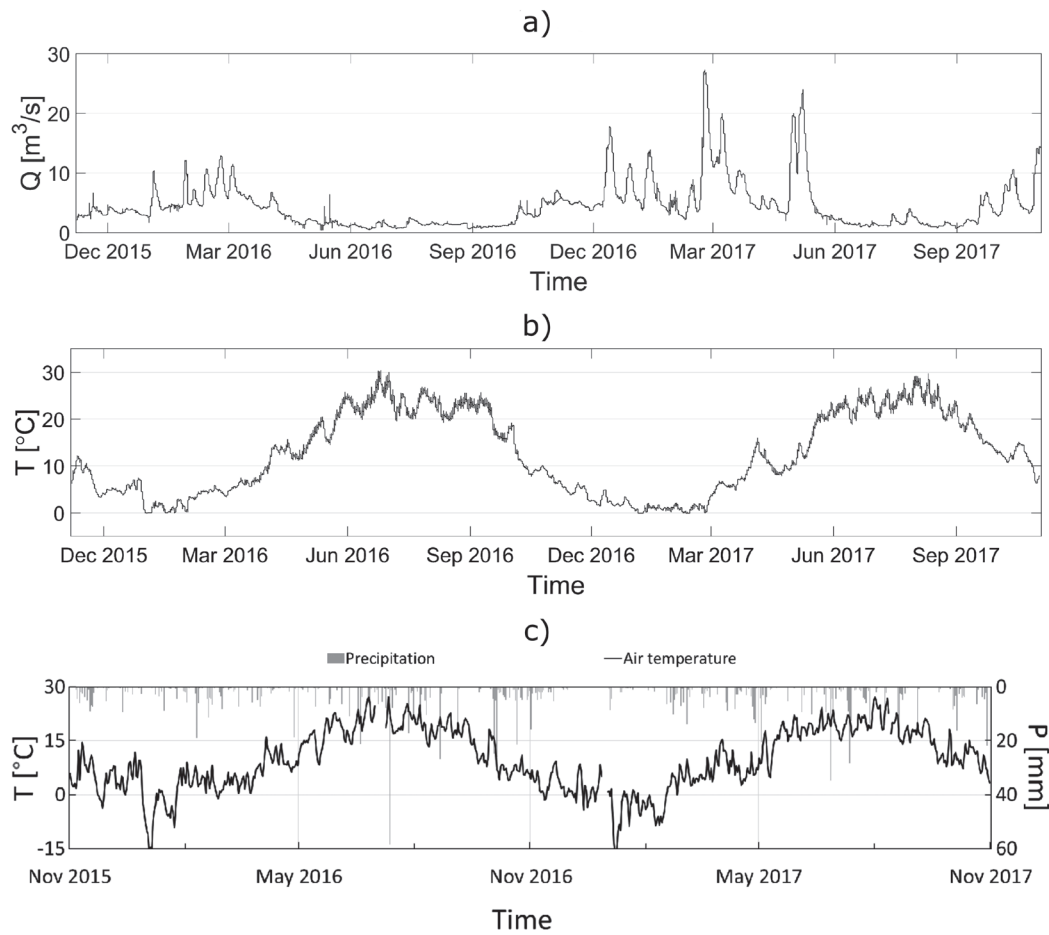


Fig. 4. Time series showing the following: the discharge Q over a 2-year period in the Świder River a), the temperature T over a 2-year period in the Vistula River b), and the daily mean air temperature T and precipitation P over a 2-year period in the Świder c).

In order to better understand the phenomenon of changes in plant biomechanics we analysed meteorological and hydrological data. Due to the lack of a gauging station on the Wilga, continuous hydrological data sets are not available. However, located close to the Wilga is the Świder River, which has very similar hydrological and morphological characteristics. An analysis of the hydrological data from gauges located in the Vistula and Świder Rivers (Fig. 4a-b) and supplementary meteorological data (Fig. 4c) allows for estimating the hydrological regime of the Wilga over a 2-year period of measurements. In addition, the water temperature, dissolved oxygen (DO), pH, and specific conductance (SPC) on the day of plant collection were measured 'in-situ' in the second season with the use of a Professional Plus (Pro Plus) multiparameter instrument (YSI, Yellow Springs, USA). The time series of the daily mean air temperature and precipitation from the meteorological station in the Świder were also investigated. Furthermore, the bathymetry and velocity spatial distribution of the Wilga in the cross-section of the sampling site were obtained with use of an acoustic Doppler current profiler (ADCP), RiverSurveyor S5 model (manufactured by SonTek, San Diego, USA).

Equipment and Plant Preparation

Transportation of plants involved their temporary placement in a 52-L plastic aquarium equipped with an aeration device. While in the laboratory, plants were stored in a 112-L tank. Natural conditions were mimicked by inclusion of two water pumps, a standard aeration device, and a fluorescent lamp with two light bulbs that were illuminated up to 12 h per day. During each measurement campaign, the photographs of individual plants and their cross-sectional areas were taken prior to biomechanical tests to define the morphology characteristics and internal structure of these aquatic macrophytes (Figs 5-6).

Biomechanical tests consisted of three-point bending and tension tests using a Tinius Olsen Bench Top Testing Machine, 5ST Model. The first step before testing was appropriate sample preparation. After removing individuals from the aquarium, the plant stem was cut into 7 cm pieces. The diameter of each part was measured with the use of a microscope or calliper. In tension tests, sample preparation included gluing short strips of sandpaper to the ends of the samples to prevent them from slipping from the machine clamps [25]. During measurements, the samples were submerged

in water (wet testing conditions) to avoid the drying of stems [details in 25]. Therefore, only a few samples were prepared at once to minimize the time that the plant was outside the aquatic environment.

Biomechanical Tests

The measurement results were processed with Horizon software included with the device. Three-point bending tests include the outcomes of

maximum deflection, maximum stress (force per area), and maximum force, which is the load at the given point of maximum deflection. The wet testing conditions affected the final values of the mechanical characteristics, which were reduced by the buoyancy forces. This step was not applied in tension tests due to a negligible influence on results. The calculated bending quantities included flexural rigidity, flexural modulus, and flexural strain. The calculations were made using the appropriate formulas [26-27]:

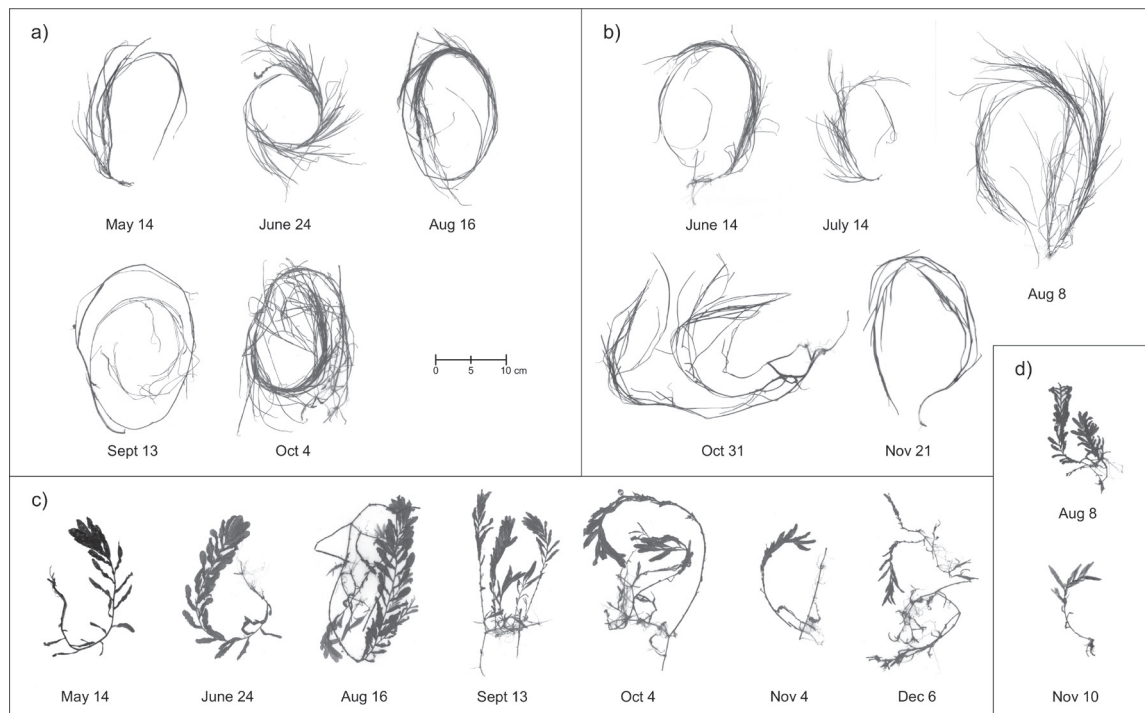


Fig. 5. Photographs of tested *P. pectinatus* L. in 2016 a) and 2017 b); and *P. crispus* L. in 2016 c) and 2017 d) at different time points.

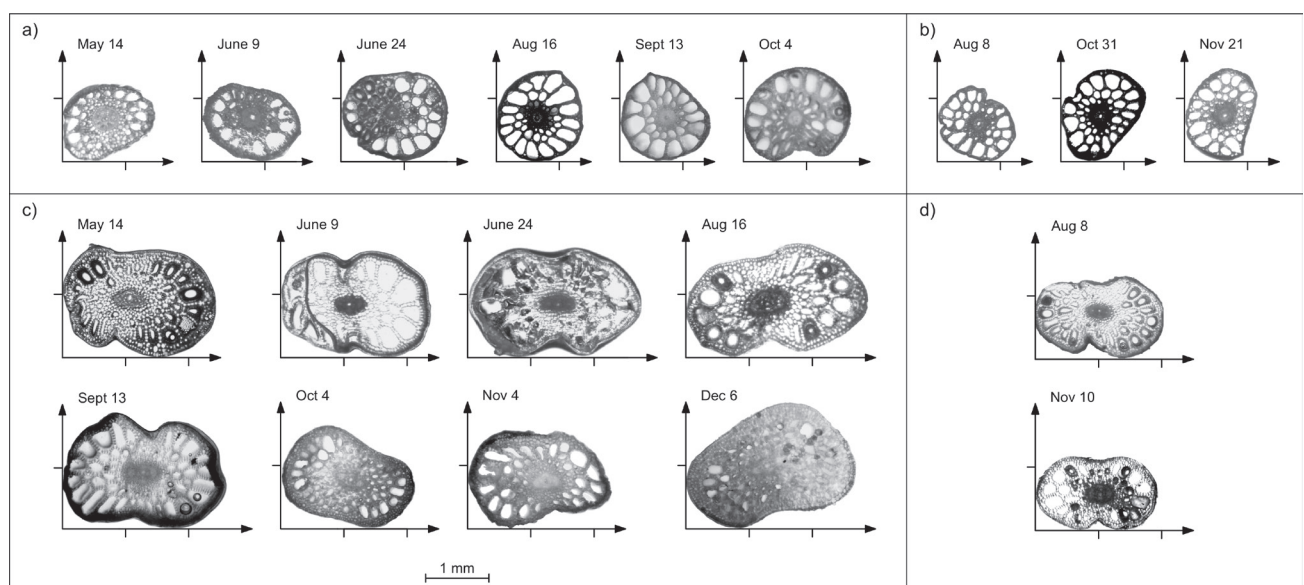


Fig. 6. Cross-sections of *P. pectinatus* L. in 2016 a) and 2017 b); and *P. crispus* L. in 2016 c) and 2017 d) in all time periods of plant collection.

- Flexural rigidity EI : $EI = \frac{S^3 F}{48 \Delta}$, where S is the span of the stem between the support bars [mm], F describes the force at a given point on the initial linear elastic part of the force-deflection curve [N], and Δ stands for deflection [mm].
- Flexural modulus E_F : $E_F = \frac{EI}{I}$, where EI is flexural rigidity [N·mm²] and I stands for the second moment of area defined as $I_C = \frac{\pi D^4}{64}$, [mm⁴] for *P. pectinatus*, circular shape; and $I_E = \frac{\pi R^3 r}{4}$, [mm⁴] for *P. crispus*, elliptical shape (D is the cross-sectional diameter, [mm], R is the long radius, and r is the short radius of the ellipse cross-section [mm]).
- Flexural strain ε_f : $\varepsilon_f = \frac{6 \Delta_{max} D}{S^2}$, where Δ_{max} is maximum deflection [mm], D is the diameter of the stem, and S stands for the span of the stem between the support bars [mm]. Tension tests were performed until the complete failure of a specimen. The Horizon software recorded data of breaking values of force, stress, and strain (maximum values at the rupture of the stem). These characteristics were needed to calculate the modulus of elasticity, i.e. Young's modulus, using formula [26]:
- Young's modulus E_T : $E_T = \frac{\sigma}{\varepsilon}$, where σ is the tensile stress [MPa] and ε stands for the strain [-].

Statistical Analysis

The bending outcomes of plant biomechanics, according to seasonal changes, were investigated through maximum force, flexural rigidity, flexural modulus, and maximum deflection, and the tensile characteristics, i.e., breaking force and Young's modulus. To analyse the differences in mean values of the biomechanical parameters between each period of plant measurements, a statistical analysis was performed using one of the resampling tests, namely the permutation test [28]. Moreover, the analysis of biomechanical traits between similar periods in seasons was also performed. For the null hypothesis, it was assumed that the mean equality contrary to the alternative hypothesis of significant differences in means between two groups of data describing an individual period of measurements. Then, the average difference module in both datasets was counted. Moreover, to find the significance of differences, two additional steps were carried out, the random mixing of values in both groups of data, keeping their numbers, and the calculation of the amount of difference for the new datasets. Further, to understand which values could be accepted when the two datasets did not differ, it was necessary to repeat the above steps. The number of 100,000 groups was considered due to the large amount of total groups. To calculate the p-value, the percentage of mean differences for random fluctuations greater than observed was taken. The differences between groups of data are statistically significant when the p-value is lower than 0.05.

Results

Hydrological and Meteorological Data

In 2016, the water temperature ranged from 2.3°C on 6 December to 24.0°C on 24 June (Table 1). The measured water discharge was at a maximum (3.62 m³·s⁻¹) on 4 November, while the minimum value (0.62 m³·s⁻¹) was observed on 13 September (Table 1). In comparison to the first season, in 2017, the water temperature values varied from 4.5°C on 21 November to 23.5°C on 12 July (Table 1). The maximum water discharge (6.68 m³·s⁻¹) was observed on 31 October, whereas the minimum value (0.76 m³·s⁻¹) was measured on 12 July (Table 1). Moreover, during the second season, the water quality parameters were measured, i.e., dissolved oxygen (DO), pH, and specific conductance (SPC). The obtained outcomes were in line with those obtained within the continuous measurement done by Rajwa-Kuligiewicz et al. [29], although the DO concentration varied from 4.28 to 9.21 mg·L⁻¹ during different periods of plant collection with the extreme value of 22.54 mg·L⁻¹ associated with very low water temperature on 21 November. The pH values ranged from 7.51 to 7.80, showing slightly alkaline water, while the conductivity oscillated around 550 μS·cm⁻¹.

In relation to the annual hydrological data, in 2016, high values of discharge (Fig. 4a) were observed during the winter and early spring, whereas later in summer it stabilised at low values (approximately 0.6 m³·s⁻¹; Fig. 4a). This period was characterised by rather steady flow conditions without major changes.

Table 1. Temperature (T) and discharge (Q) for the Wilga River for all time periods of plant collection

Day of sample collection	T (°C)	Q (m ³ ·s ⁻¹)
14 May	17.2	1.01
9 June	20.0	0.63
24 June	24.1	0.97
16 Aug.	17.7	0.91
13 Sept.	20.4	0.62
4 Oct.	15.7	1.47
4 Nov.	6.0	3.62
6 Dec.	2.3	2.57
7 June	17.9	1.10
14 June	14.9	0.89
12 July	23.5	0.76
8 Aug.	16.9	0.79
31 Oct.	5.0	6.68
10 Nov.	7.9	3.12
21 Nov.	4.5	3.23

Table 2. Diameter and three-point bending test results for all periods of measurements of *P. pectinatus* and *P. crispus* in 2016.

<i>P. pectinatus</i>									
Parameter		14 May		9 June		24 June		16 Aug.	
		Mean ±S.D.	Median	Mean ±S.D.	Median	Mean ±S.D.	Median	Mean ±S.D.	Median
Number of samples		20		20		18		19	
Diameter	mm	1.30±0.31	1.21	0.97±0.23	0.88	1.37± 0.29	1.32	1.56±0.44	1.45
Maximum force	N	0.022±0.013	0.020	0.014±0.010	0.014	0.035±0.044	0.027	0.029±0.026	0.017
Maximum stress	MPa	0.019±0.015	0.012	0.023±0.020	0.020	0.021±0.013	0.018	0.013±0.007	0.014
Flexural strain	%	2.94±1.91	2.15	3.74±1.97	4.18	5.57±2.12	5.91	5.53±2.78	6.67
Max. deflection	mm	8.41±5.40	5.12	14.44±6.63	18.85	14.92±4.82	13.27	12.70±5.99	13.89
Sec. m. of area	mm ⁴	0.196±0.222	0.104	0.063±0.092	0.029	0.225±0.206	0.149	0.435±0.456	0.217
Flexural rigidity	N·mm ²	10.63±7.77	8.19	3.29±3.42	2.21	12.01±8.12	9.83	21.09±19.27	15.72
Flexural modulus	MPa	94.51±84.66	57.82	86.59±80.25	61.40	90.18±79.13	55.72	94.09±120.77	44.63
Parameter		13 Sept.		4 Oct.		4 Nov.		6 Dec.	
		Mean ±S.D.	Median	Mean ±S.D.	Median	Mean ±S.D.	Median	Mean ±S.D.	Median
Number of samples		15		19		-		-	
Diameter	mm	1.68±0.42	1.58	1.26±0.33	1.24	-	-	-	-
Maximum force	N	0.030±0.025	0.023	0.024±0.029	0.018	-	-	-	-
Maximum stress	MPa	0.013±0.008	0.012	0.019±0.016	0.016	-	-	-	-
Flexural strain	%	6.64±1.81	6.42	4.91±2.91	5.80	-	-	-	-
Max. deflection	mm	15.32±4.86	17.79	13.94±7.11	19.29	-	-	-	-
Sec. m. of area	mm ⁴	0.531±0.456	0.306	0.178±0.198	0.116	-	-	-	-
Flexural rigidity	N·mm ²	12.65±10.53	6.06	5.31±5.86	2.72	-	-	-	-
Flexural modulus	MPa	36.66±31.63	25.75	55.08±75.78	23.47	-	-	-	-
<i>P. crispus</i>									
Parameter		14 May		9 June		24 June		16 Aug.	
		Mean ±S.D.	Median	Mean ±S.D.	Median	Mean ±S.D.	Median	Mean ±S.D.	Median
Number of samples		20		20		20		20	
Diameter	mm	2.10±0.37	2.09	2.43±0.17	2.47	1.87± 0.37	1.85	1.97± 0.40	2.04
Maximum force	N	0.071±0.039	0.069	0.069±0.031	0.063	0.057±0.033	0.048	0.081±0.044	0.070
Maximum stress	MPa	0.022±0.012	0.018	0.015±0.006	0.014	0.021±0.010	0.022	0.026±0.010	0.026
Flexural strain	%	7.81±3.11	8.00	8.64±2.68	8.24	6.64±2.07	6.91	6.48±1.92	6.04
Max. deflection	mm	13.63±4.50	14.43	13.17±4.06	13.07	12.99±3.25	13.15	12.18±2.66	11.97
Sec. m. of area	mm ⁴	1.126±0.785	0.928	1.753±0.445	1.828	0.737±0.508	0.576	0.919±0.709	0.842
Flexural rigidity	N·mm ²	25.79±17.07	22.11	36.13±24.54	29.70	25.42±15.04	21.16	65.67±34.17	62.97
Flexural modulus	MPa	33.54±34.53	21.80	19.78±11.91	16.65	51.59±34.68	41.55	105.21±75.66	82.40

Table 2. Continued.

Parameter		13 Sept.		4 Oct.		4 Nov.		6 Dec.	
		Mean ±S.D.	Median	Mean ±S.D.	Median	Mean ±S.D.	Median	Mean ±S.D.	Median
Number of samples		20		19		20		20	
Diameter	mm	2.23±0.36	2.37	2.21±0.41	2.28	1.84±0.27	1.83	1.70±0.24	1.65
Maximum force	N	0.059±0.028	0.061	0.097±0.078	0.069	0.050±0.020	0.052	0.034±0.018	0.031
Maximum stress	MPa	0.014±0.006	0.016	0.023±0.011	0.019	0.019±0.007	0.019	0.015±0.007	0.014
Flexural strain	%	9.57±2.67	10.19	8.18±1.75	8.63	7.28±1.72	6.88	6.68±2.02	6.18
Max. deflection	mm	15.57±2.90	15.33	13.88±2.73	13.82	14.66±3.23	14.73	14.40±3.54	14.07
Sec. m. of area	mm ⁴	1.383±0.672	1.536	1.363±0.776	1.327	0.640±0.404	0.551	0.457±0.289	0.360
Flexural rigidity	N·mm ²	46.28±40.97	27.12	46.93±45.79	28.42	31.55±24.58	25.71	18.41±16.25	10.51
Flexural modulus	MPa	36.38±27.46	33.08	43.17±57.69	28.74	55.22±41.36	48.99	52.71±62.24	26.04

Table 3. Diameter and three-point bending test results for all periods of measurement of *P. pectinatus* and *P. crispus* in 2017

<i>P. pectinatus</i>									
Parameter		14 June		12 July		8 Aug.		31 Oct.	
		Mean ± S.D.	Median	Mean ± S.D.	Median	Mean ± S.D.	Median	Mean ± S.D.	Median
Number of samples		19		19		19		29	
Diameter	mm	1.22±0.15	1.20	0.84±0.20	0.80	1.21±0.20	1.23	1.24±0.33	1.12
Maximum force	N	0.028±0.009	0.028	0.019±0.008	0.020	0.025±0.009	0.025	0.022±0.016	0.016
Maximum stress	MPa	0.024±0.007	0.023	0.037±0.020	0.034	0.023±0.011	0.021	0.017±0.006	0.016
Flexural strain	%	5.30±0.50	5.22	3.49±1.14	3.25	5.04±0.93	4.92	4.67±1.42	4.66
Max. deflection	mm	16.14±1.51	16.04	15.32±3.69	16.22	15.41±2.34	15.84	14.03±3.19	15.21
Sec. m. of area	mm ⁴	0.119±0.081	0.102	0.035±0.053	0.020	0.124±0.093	0.112	0.182±0.255	0.077
Flexural rigidity	N·mm ²	16.53±5.29	16.44	5.93±8.26	3.74	9.37±2.94	9.70	11.52±7.82	8.70
Flexural modulus	MPa	168.50±77.21	171.86	252.97±236.87	151.00	109.72±75.91	90.76	117.56±73.94	99.57
<i>P. crispus</i>									
Parameter		21 Nov.		8 Aug.		10 Nov.			
		Mean ± S.D.	Median	Mean ± S.D.	Median	Mean ± S.D.	Median		
Number of samples				20		16			
Diameter	mm	1.17±0.23	1.12	1.90±0.40	2.01	1.65±0.26	1.62		
Maximum force	N	0.029±0.011	0.027	0.046±0.013	0.045	0.062±0.022	0.058		
Maximum stress	MPa	0.026±0.007	0.027	0.018±0.009	0.016	0.030±0.013	0.029		
Flexural strain	%	5.04±1.00	5.08	7.67±1.95	7.81	6.78±1.64	6.85		
Max. deflection	mm	15.89±1.71	15.92	15.11±3.45	16.21	15.15±2.96	15.84		
Sec. m. of area	mm ⁴	0.116±0.122	0.077	0.793±0.498	0.802	0.420±0.291	0.334		
Flexural rigidity	N·mm ²	15.54±11.09	14.26	15.54±7.44	14.59	23.37±7.69	23.85		
Flexural modulus	MPa	174.56±88.24	163.37	38.50±41.63	19.69	77.41±47.31	79.08		

Table 4. Statistical differences between the mean values of parameters from the three-point bending tests between individual days of sample collection for *P. pectinatus* and *P. crispus*.

Day of sample collection	Mean diameter (mm)				Maximum force (N)			
	<i>P. pectinatus</i>		<i>P. crispus</i>		<i>P. pectinatus</i>		<i>P. crispus</i>	
	difference	p-value	difference	p-value	difference	p-value	difference	p-value
14 May 2016 - 9 Jun 2016	0.335	0.00042	0.331	0.00097	0.008	0.03623	0.001	0.91114
9 Jun 2016 - 24 Jun 2016	0.404	0.00003	0.557	0.00001	0.021	0.00076	0.012	0.24984
24 Jun 2016 - 16 Aug 2016	0.189	0.12757	0.100	0.41475	0.006	0.68644	0.024	0.05980
16 Aug 2016 - 13 Sep 2016	0.116	0.43741	0.262	0.03520	0.001	0.92118	0.023	0.05734
13 Sep 2016 - 4 Oct 2016	0.419	0.00289	0.028	0.81985	0.005	0.58290	0.038	0.03872
4 Oct 2016 - 4 Nov 2016	-	-	0.363	0.00260	-	-	0.047	0.00449
4 Nov 2016 - 6 Dec 2016	-	-	0.145	0.08680	-	-	0.016	0.01364
14 Jun 2017 - 12 Jul 2017	0.376	0.00001	-	-	0.009	0.00218	-	-
12 Jul 2017 - 8 Aug 2017	0.373	0.00002	-	-	0.006	0.03666	-	-
8 Aug 2017 - 31 Oct 2017	0.030	0.72896	-	-	0.003	0.49141	-	-
8 Aug 2017 - 10 Nov 2017	-	-	0.253	0.03595	-	-	0.016	0.00979
31 Oct 2017 - 21 Nov 2017	0.073	0.35038	-	-	0.007	0.07834	-	-
Day of sample collection	Flexural rigidity (N·mm ²)				Flexural modulus (MPa)			
	<i>P. pectinatus</i>		<i>P. crispus</i>		<i>P. pectinatus</i>		<i>P. crispus</i>	
	difference	p-value	difference	p-value	difference	p-value	difference	p-value
14 May 2016 - 9 Jun 2016	7.336	0.00025	10.342	0.12917	7.928	0.77013	13.759	0.10204
9 Jun 2016 - 24 Jun 2016	8.722	0.00003	10.714	0.10572	3.597	0.89382	31.804	0.00022
24 Jun 2016 - 16 Aug 2016	9.076	0.06359	40.250	0.00002	3.912	0.91184	53.618	0.00468
16 Aug 2016 - 13 Sep 2016	8.443	0.13952	19.394	0.11420	57.437	0.07082	68.824	0.00012
13 Sep 2016 - 4 Oct 2016	7.339	0.01317	0.657	0.96419	18.424	0.41336	6.784	0.77089
4 Oct 2016 - 4 Nov 2016	-	-	15.386	0.20403	-	-	12.057	0.49301
4 Nov 2016 - 6 Dec 2016	-	-	13.141	0.04592	-	-	2.513	0.89442
14 Jun 2017 - 12 Jul 2017	10.604	0.00002	-	-	84.470	0.15465	-	-
12 Jul 2017 - 8 Aug 2017	3.446	0.07533	-	-	143.253	0.01178	-	-
8 Aug 2017 - 31 Oct 2017	2.148	0.27825	-	-	7.842	0.72371	-	-
8 Aug 2017 - 10 Nov 2017	-	-	7.832	0.00354	-	-	38.907	0.01170
31 Oct 2017 - 21 Nov 2017	4.018	0.12331	-	-	57.002	0.01191	-	-
Day of sample collection	Maximum deflection (mm)				Maximum deflection (mm)			
	<i>P. pectinatus</i>		<i>P. crispus</i>		<i>P. pectinatus</i>		<i>P. crispus</i>	
	difference	p-value	difference	p-value	difference	p-value	difference	p-value
14 May 2016 - 9 Jun 2016	7.336	0.00025	10.342	0.12917	7.928	0.77013	13.759	0.10204
9 Jun 2016 - 24 Jun 2016	8.722	0.00003	10.714	0.10572	3.597	0.89382	31.804	0.00022
24 Jun 2016 - 16 Aug 2016	9.076	0.06359	40.250	0.00002	3.912	0.91184	53.618	0.00468
16 Aug 2016 - 13 Sep 2016	8.443	0.13952	19.394	0.11420	57.437	0.07082	68.824	0.00012
13 Sep 2016 - 4 Oct 2016	7.339	0.01317	0.657	0.96419	18.424	0.41336	6.784	0.77089
4 Oct 2016 - 4 Nov 2016	-	-	15.386	0.20403	-	-	12.057	0.49301
4 Nov 2016 - 6 Dec 2016	-	-	13.141	0.04592	-	-	2.513	0.89442
14 Jun 2017 - 12 Jul 2017	10.604	0.00002	-	-	84.470	0.15465	-	-
12 Jul 2017 - 8 Aug 2017	3.446	0.07533	-	-	143.253	0.01178	-	-
8 Aug 2017 - 31 Oct 2017	2.148	0.27825	-	-	7.842	0.72371	-	-
8 Aug 2017 - 10 Nov 2017	-	-	7.832	0.00354	-	-	38.907	0.01170
31 Oct 2017 - 21 Nov 2017	4.018	0.12331	-	-	57.002	0.01191	-	-

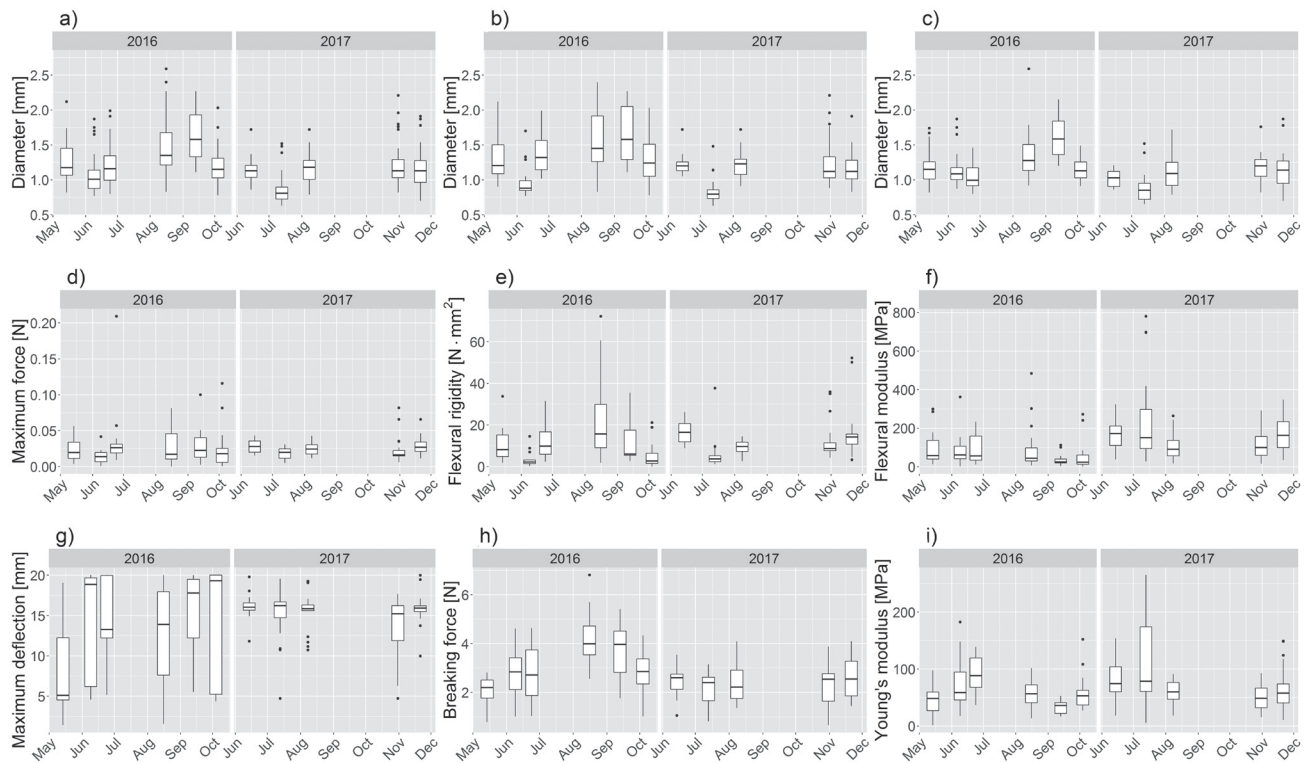


Fig. 7. Diameter of *P. pectinatus* stems of all specimens from both tests a), specimens from three-point bending tests b), specimens from tension tests c). Biomechanical parameters: maximum force d), flexural rigidity e), flexural modulus f), maximum deflection g) from three-point bending tests of *P. pectinatus* stems; breaking force h), Young's modulus i) from tension tests of plant stems on different days of sample collection in both seasons. The line in the middle of the box is the median, the box edges are the first and third quartiles, the whiskers are minimum and maximum values while the small circles represent outliers.

An increase in discharge was observed again at the beginning of autumn, whereas in winter local flooding is easily recognised. On the other hand, in 2017 the fluctuations in discharge were more frequent and the values were higher (Fig. 4a). During winter and after the spring melt period we observed flooding conditions (Fig. 4a). Later in summer, it stabilised at low values as in the previous year and again the increase began in the autumn (Fig. 4a). In comparison to discharge data, the water temperature was similar in both seasons (Fig. 4b), which was in line with the air temperature (Fig. 4c). In addition, the hydrological and meteorological data, specifically the water and air temperatures, showed that between these two years there was a shift toward warmer temperatures that started in the 2017 growing season (Fig. 4b-c).

Morphology of Plants

P. pectinatus is characterised by thinner stems in comparison to *P. crispus*. The samples were chosen randomly, and hence the means of cross-section dimensions differed between both mechanical tests. In 2016 the mean diameter of all *P. pectinatus* specimens in the three-point bending tests ranged from 0.97 to 1.68 mm in various periods of measurements (Table 2), whereas in 2017 this parameter varied from

0.84 to 1.24 mm (Table 3). The statistically significant differences confirm the decrease in diameter, which can be observed between 14 May and 9 June 2016, and 13 September and 4 October 2016, as well as between 14 June and 12 July 2017 (p-value < 0.05, Table 4, Fig. 7b). A substantial increase occurred between 9 June and 24 June 2016, and 12 July and 8 August 2017 (p-value < 0.05, Table 4, Fig. 7b). In tension tests, these values were between 1.05 and 1.60 mm in 2016 (Table 6) and between 0.89 and 1.33 mm in 2017 (Table 7), with significant increases between 24 June and 16 August 2016, 16 August and 13 September 2016, and 12 July and 8 August 2017 (p-value < 0.05, Table 5, Fig. 7c). The decreases occurred between 13 September and 4 October 2016, and between 14 June and 12 July 2017 (p-value < 0.05, Table 5, Fig. 7c). For *P. crispus*, the mean values of diameter in the three-point bending tests varied from 1.70 to 2.43 mm during the first season (Table 2) and from 1.65 to 1.90 mm during the second season (Table 3). The changes between 14 May and 9 June 2016, and 16 August and 13 September 2016 can be considered statistically significant increases, whereas between 9 June and 24 June 2016, 4 October and 4 November 2016, and 8 August and 10 November 2017 there was a significant decrease (p-value < 0.05, Table 4, Fig. 8b). The stem diameters in the tension tests ranged from 1.70 to 2.12 mm in different sampling periods

Table 5. Statistical differences between the mean values of parameters from tension tests between individual days of samples collection for *P. pectinatus* and *P. crispus*.

Day of sample collection	Mean diameter (mm)						Breaking force (N)						Young's modulus (MPa)					
	<i>P. pectinatus</i>			<i>P. crispus</i>			<i>P. pectinatus</i>			<i>P. crispus</i>			<i>P. pectinatus</i>			<i>P. crispus</i>		
	difference	p-value		difference	p-value		difference	p-value		difference	p-value		difference	p-value		difference	p-value	
14 May 2016 - 9 Jun 2016	0.018	0.84042		0.075	0.51078		0.775	0.01057		1.400	0.00137		27.836	0.02000		2.646	0.52661	
9 Jun 2016 - 24 Jun 2016	0.125	0.10652		0.140	0.31740		0.072	0.83900		0.621	0.13336		15.901	0.20174		2.386	0.59369	
24 Jun 2016 - 16 Aug 2016	0.335	0.00013		0.241	0.08060		1.440	0.00012		0.105	0.69322		36.745	0.00021		5.233	0.18888	
16 Aug 2016 - 13 Sep 2016	0.217	0.04961		0.054	0.62325		0.423	0.24186		0.564	0.28373		21.858	0.00125		9.427	0.02770	
13 Sep 2016 - 4 Oct 2016	0.449	0.00001		0.360	0.00013		0.951	0.00612		0.496	0.59070		24.461	0.00067		0.466	0.88573	
4 Oct 2016 - 4 Nov 2016	-	-		0.371	0.00030		-	-		1.599	0.04188		-	-		0.784	0.79333	
4 Nov 2016 - 6 Dec 2016	-	-		0.001	0.99176		-	-		0.177	0.65821		-	-		1.887	0.55788	
14 Jun 2017 - 12 Jul 2017	0.129	0.04386		-	-		0.277	0.20128		-	-		34.056	0.12172		-	-	
12 Jul 2017 - 8 Aug 2017	0.212	0.01535		-	-		0.217	0.42931		-	-		56.178	0.00966		-	-	
8 Aug 2017 - 31 Oct 2017	0.065	0.34034		-	-		0.202	0.49129		-	-		9.455	0.19413		-	-	
8 Aug 2017 - 10 Nov 2017	-	-		0.268	0.01023		-	-		0.147	0.64830		-	-		3.380	0.36018	
31 Oct 2017 - 21 Nov 2017	0.035	0.60056		-	-		0.414	0.09181		-	-		13.545	0.12309		-	-	

Table 6. Diameter and tension test results for all periods of measurement of *P. pectinatus* L. and *P. crispus* L. in 2016.

<i>P. pectinatus</i>									
Parameter		14 May		9 June		24 June		16 Aug.	
		Mean±S.D.	Median	Mean±S.D.	Median	Mean±S.D.	Median	Mean±S.D.	Median
Number of samples		20		20		20		20	
Diameter	mm	1.19±0.25	1.15	1.17±0.28	1.09	1.05±0.19	1.00	1.38± 0.37	1.28
Breaking force	N	2.05±0.59	2.20	2.82±1.15	2.84	2.75±1.09	2.71	4.19±1.03	3.99
Breaking stress	MPa	2.07±0.93	2.17	2.81±1.34	2.66	3.30±1.25	3.48	3.01±0.87	3.25
Breaking strain	%	7.04±1.88	6.52	7.48±2.29	7.84	6.22±2.10	5.65	11.13±2.30	11.34
Young's modulus	MPa	47.79±26.58	48.42	75.62±44.33	58.51	91.52±32.25	88.25	54.78±23.61	56.48
Parameter Mean±S.D.		13 Sept.		4 Oct.		4 Nov.		6 Dec.	
		Mean±S.D.	Median	Mean±S.D.	Median	Mean±S.D.	Median	Mean±S.D.	Median
Number of samples		18		20		-		-	
Diameter	mm	1.60± 0.28	1.59	1.15±0.16	1.13	-	-	-	-
Breaking force	N	3.77±1.17	3.96	2.81±0.80	2.86	-	-	-	-
Breaking stress	MPa	1.86±0.36	1.85	2.75±0.82	2.68	-	-	-	-
Breaking strain	%	10.31±1.82	10.38	8.05±1.48	8.11	-	-	-	-
Young's modulus	MPa	32.92±11.60	36.05	57.38±30.23	52.83	-	-	-	-
<i>P. crispus</i>									
Parameter Mean±S.D.		14 May		9 June		24 June		16 Aug.	
		Mean±S.D.	Median	Mean±S.D.	Median	Mean±S.D.	Median	Mean±S.D.	Median
Number of samples		20		19		19		19	
Diameter	mm	2.01±0.30	2.04	2.08±0.40	2.09	1.94± 0.45	1.78	1.70± 0.38	1.77
Breaking force	N	2.44±0.84	2.26	3.84±1.58	3.66	3.22±0.76	3.21	3.11±0.87	3.18
Breaking stress	MPa	0.80±0.31	0.70	1.17±0.52	1.03	1.20±0.47	1.08	1.61±0.92	1.37
Breaking strain	%	7.77±3.27	7.66	6.68±1.89	5.94	7.79±2.48	7.61	7.73±3.14	7.25
Young's modulus	MPa	16.34±7.72	15.26	18.98±15.63	12.98	21.37±10.60	19.08	26.60±13.35	25.59
Parameter Mean±S.D.		13 Sept.		4 Oct.		4 Nov.		6 Dec.	
		Mean±S.D.	Median	Mean±S.D.	Median	Mean±S.D.	Median	Mean±S.D.	Median
Number of samples		17		22		19		20	
Diameter	mm	1.76±0.26	1.77	2.12±0.27	2.03	1.75±0.34	1.78	1.74±0.28	1.70
Breaking force	N	3.68±1.96	3.15	4.17±3.28	2.99	2.57±0.64	2.53	2.40±1.46	1.74
Breaking stress	MPa	1.46±0.55	1.29	1.12±0.74	0.91	1.17±0.49	0.98	1.03±0.55	0.89
Breaking strain	%	8.97±2.56	8.06	7.42±2.70	7.75	8.39±2.50	8.72	6.85±2.65	6.04
Young's modulus	MPa	17.18±10.91	15.59	17.64±8.34	16.84	16.86±10.40	14.11	18.75±9.48	16.97

in 2016 (Table 6), while in 2017 the values obtained were between 1.44 and 1.70 mm (Table 7). The calculations show that between 13 September and 4 October 2016 there was a significant increase, while a substantial decrease was observed between 4 October and 4 November 2016, and 8 August and 10 November 2017 (p-value < 0.05, Table 5, Fig. 8c).

The two species tested from the family *Potamogetonaceae* differ in structure (Figs 5 and 6).

The cross-sectional area of *P. pectinatus* has a circular shape, while in *P. crispus*, the form resembles an ellipse (Fig. 6). However, both species have a honeycomb aerenchyma structure (Fig. 6). In the first season in 2016, *P. pectinatus* was most developed in September, but the plant was characterised by a lower number of leaves (Fig. 5). Additionally, the time when *P. crispus* was in full bloom is difficult to estimate due to its unique life cycle [20] (Fig. 5).

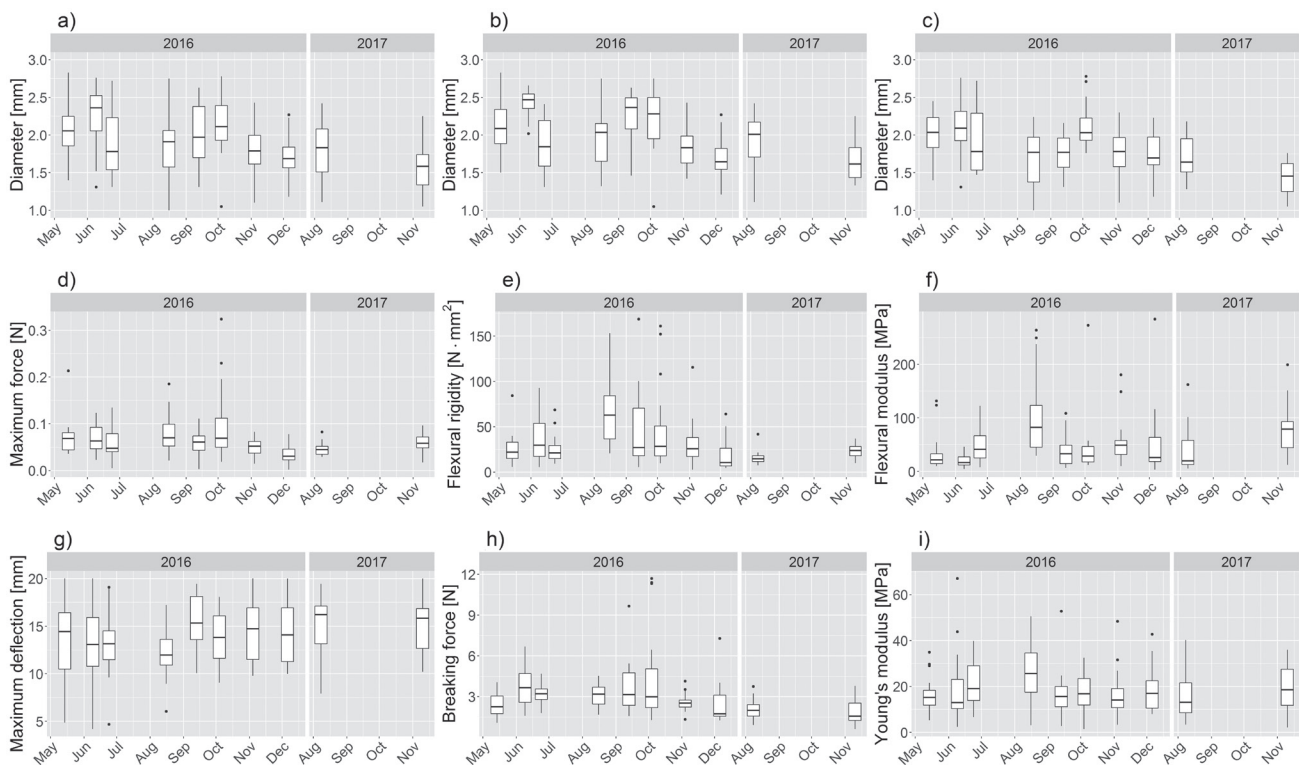


Fig. 8. Diameter of *P. crispus* stems of all specimens from both tests a), specimens from three-point bending tests b), specimens from tension tests c). Biomechanical parameters: maximum force d), flexural rigidity e), flexural modulus f), maximum deflection g) from three-point bending tests of *P. crispus* stems; breaking force h), Young's modulus i) from tension tests of plant stems on different days of sample collection in both seasons. The line in the middle of the box is the median, the box edges are the first and third quartiles, the whiskers are minimum and maximum values while the small circles represent outliers.

Table 7. Diameter and tension test results for all periods of measurements of *P. pectinatus* and *P. crispus* in 2017.

<i>P. pectinatus</i>									
Parameter		14 June		12 July		8 Aug.		31 Oct.	
		Mean±S.D.	Median	Mean±S.D.	Median	Mean±S.D.	Median	Mean±S.D.	Median
Number of samples		19		17		19		25	
Diameter	mm	1.02±0.12	1.03	0.89±0.24	0.85	1.10±0.24	1.09	1.17±0.19	1.20
Breaking force	N	2.47±0.60	2.60	2.19±0.68	2.40	2.41±0.88	2.22	2.20±0.94	2.54
Breaking stress	MPa	3.06±0.87	2.93	4.12±2.24	3.82	2.52±0.50	2.68	2.08±0.81	2.23
Breaking strain	%	7.04±1.99	7.71	7.93±2.87	7.88	9.21±2.65	9.40	7.23±2.39	7.92
Young's modulus	MPa	82.18±38.57	74.33	116.23±83.72	78.71	60.05±21.61	59.76	50.60±23.86	48.52
<i>P. crispus</i>									
Parameter Mean±S.D.		21 Nov.		8 Aug.		10 Nov.			
		Mean±S.D.	Median	Mean±S.D.	Median	Mean±S.D.	Median		
Number of samples		27		19		12			
Diameter	mm	1.33±0.27	1.13	1.70±0.28	1.64	1.44±0.24	1.46		
Breaking force	N	2.62±0.79	2.55	2.06±0.72	1.99	1.91±1.07	1.56		
Breaking stress	MPa	2.72±0.82	2.60	0.95±0.44	0.80	1.27±0.83	0.99		
Breaking strain	%	8.52±2.38	8.79	7.02±2.61	6.65	6.65±2.30	6.90		
Young's modulus	MPa	64.14±36.56	57.83	15.75±9.37	13.13	19.13±10.59	18.57		

Comparison between Seasons

The differences in diameters from three-point bending and tension tests, flexural rigidity, and Young's modulus between similar measurement periods of the first and second seasons were investigated. The results for the three-point bending tests of *P. pectinatus* showed that there were significant differences in diameter between 9 June 2016 and 14 June 2017, 24 June 2016 and 12 July 2017, and 16 August 2016 and 8 August 2017 (p -value<0.05, Table 8). As for flexural rigidity, significant changes were observed in all investigated periods (p -value<0.05, Table 8). In comparison to the three-point bending tests, in tension tests the only significant difference occurred in diameter between 16 August 2016 and 8 August 2017 (p -value<0.05, Table 8). Young's modulus values did not change significantly (Table 8). For *P. crispus*, despite no changes in diameter between 16 August 2016 and 8 August 2017 in both tests (p -value<0.05, Table 8), the comparison showed significant differences in biomechanical parameters, i.e., flexural rigidity and Young's modulus (p -value<0.05, Table 8). On the other hand, the statistical analysis of these parameters between 4 November 2016 and 10 November 2017 showed significant differences in diameters (p -value<0.05, Table 8); however, for flexural rigidity and Young's modulus, there were no changes.

Three-Point Bending Tests

In 2016, at the beginning of the investigated period between 14 May and 9 June, the maximum force and the flexural rigidity of *P. pectinatus* significantly decreased (p -value < 0.05, Table 4, Fig. 7d-e). Then, those values rose rapidly between 9 June and 24 June, which was confirmed by statistical analysis (p -value<0.05, Tables 2 and 4, Fig. 7d-e). From the end of June to October, the variations in force outcomes were not statistically significant. Despite this phenomenon, changes in the mechanics of *P. pectinatus* were observed. The significant drop in the flexural rigidity in October (p -value < 0.05, Table 4, Fig. 7e) occurred after a period when the plant was characterised by the highest stiffness, with mean values in August and September equalling 21.09 N·mm² and 12.65 N·mm², respectively (Table 2). The statistical analysis has shown that there were no significant changes in the flexural modulus between the investigated periods. However, there was a visible downward trend with passing time (Table 2, Fig. 7f). During the next season in 2017, the statistical analysis showed a decrease in maximum force and flexural rigidity between 14 June and 12 July (p -value<0.05, Table 4, Fig. 7d-e). Then, the maximum force significantly increased between 12 July and 8 August (p -value < 0.05, Table 4, Fig. 7d). At the same time, there was observed drop in the flexural modulus, while an increase occurred between 31 October and 21 November (p -value < 0.05, Table 4, Fig. 7f). Significant

changes in maximum deflection were observed between October 31 and November 21 (p -value<0.05, Table 4, Fig. 7g).

On the other hand, in 2016 *P. crispus* was characterised by a substantially increasing flexural modulus that mimicked plant growth. The noticeable increase in this parameter was observed between 9 June and 24 June, and 24 June and 16 August (p -value<0.05, Table 4, Fig. 8f), when it reached a mean value of 105.21 MPa (Table 2). After this period, the modulus of elasticity significantly decreased to 36.38 MPa (p -value<0.05, Tables 2 and 4, Fig. 8f). In contrast to *P. pectinatus*, the maximum bending force of *P. crispus* was balanced during the warmer months (May–September) and subsequently increased in October (p -value<0.05, Tables 2 and 4, Fig. 8d). After that, a significant decrease in bending strength was observed to the end of the investigated period (p -value < 0.05, Table 4, Fig. 8d). For flexural rigidity, statistically significant changes (p -value < 0.05, Table 2, Fig. 8e) were obtained between 24 June and 16 August, and between 4 November and 6 December. Like *P. pectinatus*, the highest mean value was observed in August, equalling 65.67 N·mm² (Table 2). In the second season, this species was found in exactly the same place twice, and statistical calculations between periods (8 August and 10 November) showed significant increases in maximum force, flexural rigidity, and flexural modulus (p -value<0.05, Table 4, Fig. 8d-f).

In a comparison of both hydrophyte species, differences in outcomes of the three-point bending parameters were visible. The forces needed to completely fracture *P. crispus* were much higher than those needed to fracture *P. pectinatus*. A direct reason could be the greater stiffness resulting from the thickness of shoots, which was two to ten times greater than in *P. crispus* (Table 2). In contrast to flexural rigidity, a larger resistance to being deformed elastically was obtained for *P. pectinatus* due to its thinner shoots (Table 2).

Tension Tests

During the first investigated season in 2016, statistically significant variations in mean values of breaking forces for *P. pectinatus* L. were observed between 14 May and 9 June, 24 June and 16 August, and 13 September and 4 October (p -value<0.05, Table 5, Fig. 7h). There was an increase in plant strength until August, with a maximum value of 4.19 N (Table 6), whereas at the end of the vegetative season there was visible drop in values to 2.81 N on the 4 October (Table 6). In the period between May and June, the shoot dimensions were similar, and no significant changes were noted (p -value<0.05, Table 5, Fig. 7c). Nevertheless, Young's modulus increased sharply from 47.79 up to a maximum value of 91.52 MPa (Table 6). Furthermore, even with increasing plant dimensions, shoot elasticity drastically decreased to 32.92 MPa

Table 8. Statistical differences in the mean values of the diameter and flexural rigidity from the three-point bending test; the diameter and Young's modulus from the tension tests between similar measurement periods from the first and second seasons for *P. pectinatus* and *P. crispus*.

Day of sample collection	Three-point bending test				Tension test			
	Diameter (mm)		Flexural rigidity (N·mm ²)		Diameter (mm)		Young's modulus (MPa)	
	difference	p-value	difference	p-value	difference	p-value	difference	p-value
<i>P. pectinatus</i>								
9 Jun 2016 - 14 Jun 2017	0.25	0.00023	13.240	0.00001	0.153	0.02844	6.554	0.62476
24 Jun 2016 - 12 Jul 2017	0.532	0.00001	6.086	0.02746	0.157	0.03287	24.710	0.23322
16 Aug 2016 - 8 Aug 2017	0.348	0.00308	11.716	0.00592	0.281	0.00723	5.277	0.48248
4 Oct 2016 - 31 Oct 2017	0.015	0.88269	6.214	0.00333	0.017	0.75732	6.781	0.41406
<i>P. crispus</i>								
16 Aug 2016 - 8 Aug 2017	0.068	0.59424	50.127	0.00001	0.068	0.59424	50.127	0.00001
4 Nov 2016 - 10 Nov 2017	0.191	0.04179	8.173	0.22460	0.311	0.01147	2.275	0.56375

(p-value 0.05, Tables 5-6, Fig. 7i). An increase in Young's modulus was observed between September 13 and October 4 (p-value 0.05, Table 5, Fig. 7i). In the second season of 2017, the breaking force of *P. pectinatus* did not change significantly (Table 5); however, there a statistically significant decrease in Young's modulus was observed between 12 July and 8 August (p-value 0.05, Table 5, Fig. 7i). Generally, in various phases of plant growth the results confirm that the biomechanical properties of stems vary according to seasonal changes.

In contrast to *P. pectinatus*, *P. crispus* had more stable properties. An increase in mean breaking force values during the first season was observed between 14 May and 9 June (p-value 0.05, Tables 5-6, Fig. 8h), when the forces increased from 2.44 to 3.84 N (Table 6). During the summer period, the fluctuations were registered as irrelevant. However, when the water temperature drastically dropped (to 6°C on the 4 November, Table 1), a statistically significant decrease in tensile load of approximately 40% (p-value 0.05, Tables 5-6, Fig. 8h) was observed. The parameter of elasticity of the plant stem, Young's modulus, remained at a similar level until August. Then, the ability to elastically deform was reduced to 17.18 MPa (p-value 0.05, Tables 5-6, Fig. 8i), and it oscillated within 18 MPa (Table 6). In the second season, there were no significant changes in biomechanical properties for this species (Table 5).

Discussion

The biomechanical behaviour of plants is strongly related to flow-vegetation-sediment interactions [30], which is important for engineering and flood protection. The presented data and calculations based on the biomechanical tests, i.e., three-point bending and tension, showed that fluctuations in mechanical

traits occurred in both tested plant species during the investigated seasons. The variations may be a consequence of many factors, e.g., plant growth stage or hydrological and meteorological conditions.

As previously mentioned, *Potamogeton* spp. were only investigated biomechanically by Brewer and Parker [22], Bociąg et al. [23], and Miler et al. [16]. For instance, *P. pectinatus* was examined for the first time by Brewer and Parker [22]. The plant was subjected to tensile tests; however, the values for tensile strength (33.8 MPa) differed significantly from those we have obtained in all periods of our study (0.80-3.30 MPa, Table 6). Bociąg et al. [23] collected individual plants from stagnant and flowing water in July. They found values of breaking force (3.3 N) for plants coming from rivers that were more comparable to our outcomes around the same time of the year (Table 6). The second species, *P. crispus*, was investigated by Miler et al. [16]. They obtained lower values of biomechanical traits, i.e., breaking force, Young's modulus, than our results, which may be related to a smaller sample diameter. However, the flexural modulus was much higher than the results presented in this paper. Nevertheless, the comprehensive research on the seasonal variability of the biomechanical properties of these two species is presented for the first time here. The comparison of two seasons for *P. pectinatus* showed that there was a repeating cycle of changes in biomechanical traits (Fig. 7); however, some shifts of similar trends in characteristics have been noticed, i.e., the statistical analysis demonstrated that at the same time of year, the differences in flexural rigidity and diameters from both tests were significant (Table 8, Fig. 7). This shift may be related to changes in hydrological and meteorological conditions. The vegetation season started later in 2017 than in 2016, and it was shorter (Fig. 4b-c). We speculate that this shift is related to a clearly longer period of lower air and water temperatures (Table 1). This phenomenon indicates the need to take a broader perspective in determining

the biomechanical parameters of a plant from a particular period, e.g., by considering air and water temperatures. According to O'Hare [12], aquatic macrophytes may shift their adjustments to the conditions, and they may become more stress-tolerant. This is confirmed by the differences in the results of the three-point bending test between seasons, where in 2017 plants were characterised by lower flexural rigidity connected with higher flexural modulus, which was responsible for elastic deformation, meaning the return of the plant to its original shape after the load was removed. The higher discharges and water velocities could cause the adoption of an avoidance strategy to prevent damage or destruction, which was characterised by an increase in shoot elasticity in line with the results of the study by Robioneck et al. [31].

According to Schutten et al. [32] and Puijalón et al. [33], aquatic plants are flexible and adjust to living factors by bending with the flow direction. This is an example of an avoidance strategy that helps them survive in different hydrological conditions. This statement is confirmed by our results as the *Potamogeton* spp. requires a higher tensile strength to avoid destruction due to the flexibility of its shoots, which bend easily (Tables 2-3, 6-7). However, the investigated hydrophytes, which have different structures and life cycles, show significant differences in biomechanical properties. Moreover, the flow velocities in the cross-section at the location of the plant habitats differed for both species. *P. pectinatus* grew in a place where the flow was considerably lower than in the habitat of *P. crispus* (Fig. 1), which suggests that higher resistance of *P. crispus* is one of the traits of the avoidance strategy adapted by this plant.

Conclusions

Our results supplement an aquatic plant biomechanics database that has been successfully built during recent years [15-16, 22-23, 32-33], complementing it by providing data on the seasonal changes of two commonly growing aquatic plants, which were, for the first time, collected during two vegetation seasons. We believe that the continuous observation and measurement of biomechanical properties of aquatic plants are required, as they will provide essential information about plant adaptive capacity to changing conditions. This is significant, since hydraulic resistance is the most important factor for ecological and civil engineering management.

Moreover, the following five points summarize our conclusions:

1. The calculations of biomechanical traits – particularly strain, stiffness, and the moduli of elasticity – clearly indicate the significant difference in biomechanical properties of two submerged aquatic plants *P. pectinatus* and *P. crispus* at the same moment in phenological time.

2. *Potamogetonaceae* plants were characterised by approximately similar strains under tension. The main difference was noticeable in the modulus of elasticity values, where *P. pectinatus* was much more prone to return to its original shape after the removal of the acting forces.
3. The differences in the structures and life cycles of the investigated species may be crucial factors that differentiate the mechanical strength to withstand bending and tensile loads of both macrophytes.
4. The hydrological and meteorological conditions may be considered the main factors regulating the values of biomechanical traits of *Potamogeton* spp., with flexural rigidity being the most sensitive parameter for changes in hydrological conditions.
5. The comparison of data of the biomechanical tests, even for the same species and at the same time of year, may lead to misinterpretation of the obtained results, as the statistical analysis demonstrated that the differences in flexural rigidity and diameters from both tests could be significant. This makes it very difficult to compare these results to earlier results obtained by other researchers, as the stage of plant growth was often omitted.

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Conflict of Interest

The authors declare no conflict of interest.

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